

Neuropathology and therapeutic management of Alzheimer's disease – an update

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Abstract

Alzheimer's disease (AD) is a progressive neurodegenerative disorder which is the principal cause of dementia throughout the world and the fourth cause of death in developed countries. The pathological hallmarks of this disease are regionalized neuronal dysfunction/death, accumulation of senile plaques extracellularly and neurofibrillary tangles (NFTs) intraneuronally. Several hypotheses have been put forth to explain the pathophysiology of this disease, including aberrant β -amyloid ($A\beta$) metabolism, hyperphosphorylation of cytoskeletal proteins, genetic predisposition (mutations in genes coding for presenilin-1 and -2 (PS-1 and PS-2) and amyloid precursor protein (APP), apolipoprotein E genotype, oxidative stress, excitotoxicity, inflammation and abnormal cell cycle re-entry. However, none of these hypotheses alone is sufficient to explain the diversity of biochemical and pathological abnormalities in AD. Currently, medications approved by the United States FDA for AD include acetylcholinesterase (AChE) inhibitors and memantine. However, these drugs provide only symptomatic relief and do not stop disease progression. The major focus of research now is to find novel therapeutic drug candidates targeting the underlying pathophysiological mechanisms. These therapeutic strategies include drugs targeting amyloid and tau pathology, immunotherapy, neurotransmitter replacement therapy, nutraceuticals and disease-modifying therapies. The main focus of this review is to provide new insight on the various mechanisms involved in the neuropathology of AD and shed light on current and future treatment strategies aimed at improving both cognitive deficits and halting the deadly neurodegenerative progression of the disease.

Introduction

Alzheimer's disease (AD) is the most common age-related neurodegenerative disorder (1). It is characterized by progressive memory loss, impairments in language and visuospatial skills, episodes of psychosis, aggressiveness and agitation, ultimately leading to death (2, 3). Histologically, it is characterized by two main structural changes in the brain: intracellular protein deposits termed neurofibrillary tangles (NFTs) and extracellular amyloid protein deposits surrounded by dystrophic neuritis that contribute to senile plaques (4) (Fig. 1).

In 1907, 100 years ago, Alois Alzheimer described two pathological alterations in the brain of a female patient suffering from dementia (5). Alzheimer had described a 'peculiar substance' occurring as extracellular deposits in specific brain regions, which are now referred to as amyloid plaques. In the mid-1980s it was discovered that the plaques consisted of aggregates of small peptides characterized as amyloid plaques (6, 7). The second lesion described by Alzheimer were the NFTs, which were later found to be composed of abnormally hyperphosphorylated tau protein (8, 9). Although

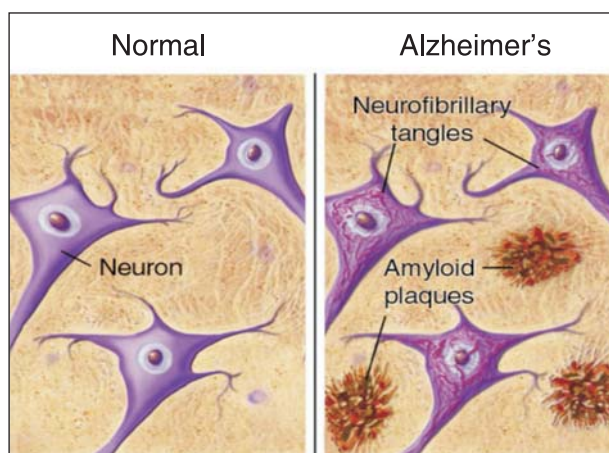


Fig. 1. A comparison of the normal and AD brain neurons where the AD brain shows the presence of the hallmark features, i.e., amyloid plaques and neurofibrillary tangles.

plaques and NFTs are the main significant pathological changes occurring in the AD brain, other structural and functional alterations also ensue, including inflammatory responses and oxidative stress (10, 11).

Epidemiological studies reveal that AD affects nearly 25 million patients worldwide, with over 4 million in the U.S. alone. As the average life expectancy increases throughout the world, it is expected that the number of people at risk of developing AD will increase significantly. In fact, it is estimated that by 2050, 14 million Americans will have AD if preventive treatment does not become available (12). This debilitating disorder poses a great financial burden not only on society but also on caregivers. The minimal cost of treatment is estimated to be upwards of USD 100 billion per year, a figure that will rise as the prevalence increases (13).

Despite these daunting statistics and the progress made in understanding the molecular and cellular basis of this disorder, there is still a need to develop an effective therapy for the management of AD. Current medications that have received FDA approval for the treatment of AD include acetylcholinesterase (AChE) inhibitors for mild to moderate cases, and memantine, an NMDA receptor antagonist for the treatment of moderate to severe Alzheimer's dementia. However, these drugs only provide symptomatic relief in some patients but do not halt disease progression (14, 15). Thus, there is an enormous medical need to develop novel therapeutic strategies that target the underlying neuropathological mechanisms involved in the progression of AD.

The aim of this review is to provide an overview of the neuropathology of AD and present and future therapeutic strategies for AD which could be employed for effective management of this progressive neurodegenerative disorder.

Etiopathogenesis of AD

The etiology and pathophysiology of AD are multifactorial and several independent hypotheses have been proposed to link the pathological lesions and the neuronal cytopathology. However, none of these theories alone is sufficient to explain the diversity of biochemical and pathological abnormalities of AD (16). It is reasonable to speculate that AD occurs as a result of a series of insults to the brain. Various factors that are involved in the etiopathogenesis of AD have been suggested.

β -Amyloid

The amyloid cascade hypothesis is the most popular hypothesis which explains the mechanisms leading to AD. It states that A β , a fragment of the amyloid precursor protein (APP) plays a central role in the pathogenesis of AD (17). It is believed that accumulation of A β (in particular the A β_{42} peptide) in the brain initiates a cascade of events that ultimately leads to neuronal dysfunction, neurodegeneration and dementia (18). More recent findings suggest that rather than highly aggregated A β species,

soluble prefibrillar forms of A β (so-called A β -derived diffusible ligands, or ADDLs) may represent the neurotoxic entity in synaptic dysfunction (19, 20).

A β is generated by proteolytic cleavage of APP, an integral membrane protein, by two proteases termed β - and γ -secretase (21). APP cleavage by α -secretase, which was the first proteolytic cleavage to be identified, precludes A β generation since the α -secretase cleavage site is located within the A β sequence (22, 23). A brief outline of APP processing by the amyloidogenic and non-amyloidogenic pathways is represented in Figure 2. The accumulation of A β is thought to play a pivotal role in neuronal loss or dysfunction through a cascade of events that include oxidative stress, mitochondrial oxidative damage and inflammatory processes (24, 25). In summary, A β remains a putative pathogenic species involved in the progression of AD (26, 27).

Hyperphosphorylated tau

Abnormally phosphorylated tau is the major constituent of NFTs. Hyperphosphorylation causes disengagement of tau from microtubules and aggregation of the filamentous protein (9). The microtubule cytoskeleton and its numerous microtubule-associated proteins (MAPs) are involved in transporting the signaling molecules at a synapse to the nucleus to induce the appropriate responses (28). MAP tau also plays an important role in the developmental maturity of neurons. Thus, disturbances and disruption of these proteins lead to synaptic and neuritic atrophy. AD is characterized by abnormal orientation and depletion of dendritic microtubules (29, 30) and abnormal hyperphosphorylated MAP tau (9).

Hyperphosphorylated tau (paired helical filaments, or PHF-tau) is unable to bind the assembled microtubules (31) and indirectly disrupts cytoskeletal integrity, leading to neuronal dysfunction and NFT formation (32, 33) (Fig. 3). However, the molecular details of tau-related neurodegeneration are still not well understood.

Oxidative stress and cell cycle dysregulation

Oxidative stress can be defined as the breaching of the oxidant defenses with consequent detrimental cellular effects (34). Free radicals that are produced during oxidative stress are speculated to be pathologically important in AD (35, 36). Modifications driven by oxidative stress have been observed in virtually all classes of biomacromolecules in association with susceptible neurons of AD. These include DNA and RNA oxidation, which is marked by increased levels of 8-hydroxyguanosine (37, 38), protein oxidation involving the oxidative modification of various enzymes involved in ATP synthesis (34, 39), lipid peroxidation (40) and modification of sugars marked by increased glycation and glycooxidation (36, 41) (Fig. 4).

There is also increasing evidence that dysregulation of the cell cycle coupled with oxidative stress in hippocampal neurons initiates the pathophysiological cascade in AD (42). This is supported by the ectopic expres-

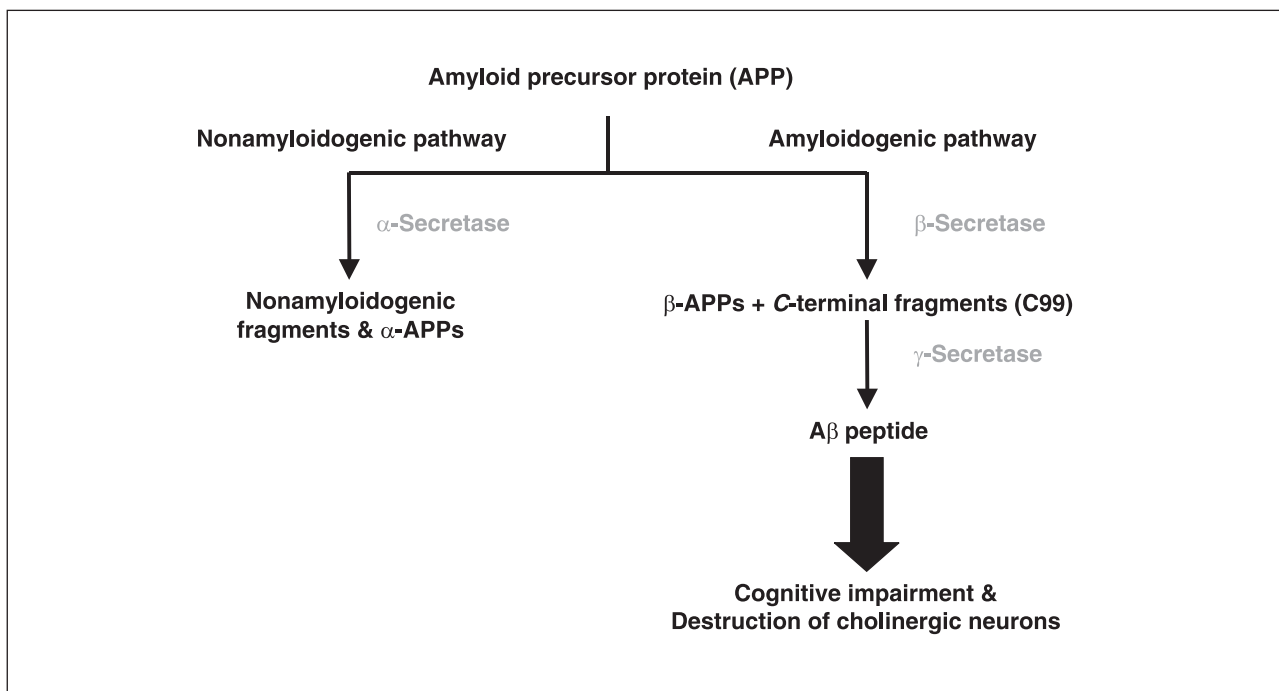


Fig. 2. Processing of amyloid precursor protein by α -, β - and γ -secretase to release β -amyloid (A β) peptide, the main culprit in AD pathophysiology.

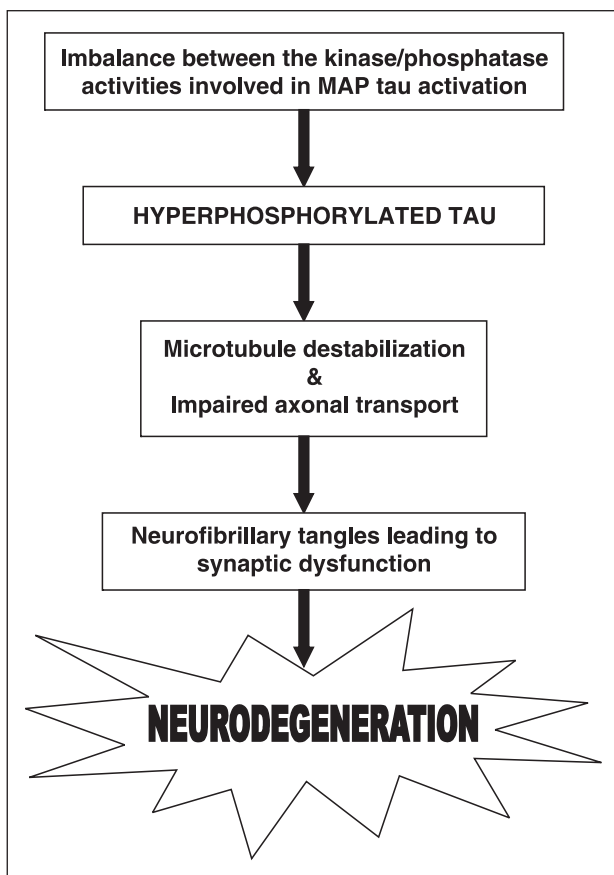


Fig. 3. Sequence of events leading to neurofibrillary tangle formation and neurodegeneration in AD.

sion of cell cycle markers (43), organelle kinesis (44) and cytoskeletal alterations. Cell cycle alterations lead to tau phosphorylation, a fact supported by the increased expression of proteins like cyclin D and cyclin-dependent kinase 4 (CDK4) involved in the G0/G1 transition during mitosis (45, 46). This cell cycle dysregulation plays an important role in the apoptotic death of postmitotic neurons (47), which finally leads to neurodegeneration. Thus, cell cycle dysregulation and oxidative stress work in synergy in the development of AD (42).

Evidence also indicates that cerebral metabolism is reduced in AD (48). The key player in this scenario is mitochondrial dysfunction (44), which results in oxidative phosphorylation and the generation of reactive oxygen species (ROS). These ROS not only have cellular targets but also damage the mitochondrial components themselves, particularly mitochondrial DNA, which is highly susceptible to oxidative stress due to a lack of histones (34). These dysfunctional mitochondria promote the interaction between redox metals and oxidative response elements (49), causing further neuronal damage.

Genetic mutations

Certain causative genetic mutations have been identified that account for less than 5% of the cases of AD, known as familial AD. These include mutations in genes coding for APP, presenilin-1 (PS-1) and presenilin-2 (PS-2). APP is a transmembrane glycoprotein that is proteolytically cleaved to generate A β (50). PS-1 is the most critical component of the γ -secretase complex and mutations in this gene cause the production of highly amy-

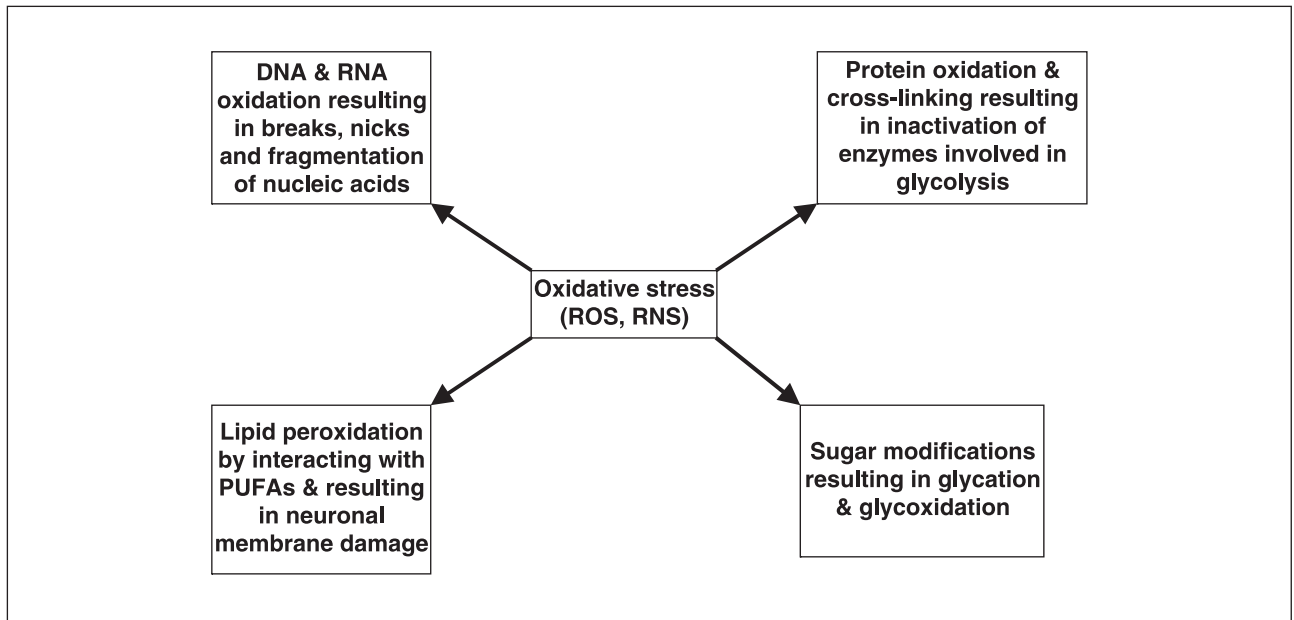


Fig. 4. Interplay between oxidative stress, nucleic acid oxidation, lipid peroxidation and protein oxidation and sugar modifications in AD.

loidogenic $A\beta_{42}$ (51-53). PS-2 is structurally similar to PS-1 but has distinct functions. It is important for the formation of a fully functional γ -secretase complex (50) and is involved in the proteolytic processing of APP. These mutations lead to increased production of $A\beta$, which aggregates in the brain to form the characteristic amyloid plaques (54, 55).

Neuroinflammation

In the AD brain, degenerating neurons, aggregated $A\beta$ protein and NFTs are sites of neuroinflammation (56). Evidence highlights the strong association between AD and abnormal activity of proinflammatory cytokines such as interleukin-1 β (IL-1 β) (57), IL-6, transforming growth factor- β (TGF- β) (58), IL-18 and TGF- β 1 (59). $A\beta$ aggregates appear to be involved in triggering reactivity of glial cells, with the consequent release of nitric oxide (NO) (60), nerve growth factor (NGF) and signaling through the p75 cell death receptor (61). Abnormal release of IL-1 and IL-6 also affects the tau phosphorylation patterns and other intracellular events linked to neuronal degeneration (62). In addition to these findings, overexpression of IL-6 in the brain of transgenic mice that overproduce cytokines (63) is associated with gliosis and disruption of cholinergic neurotransmission in the hippocampus (64). Thus, $A\beta$ itself stimulates the production of inflammatory cytokines by astrocytes and microglia, thereby triggering the early events of neurodegeneration associated with AD (65).

Aberrant lipid metabolism

Hypercholesterolemia and elevated levels of $A\beta$ are linked to a risk for AD (66, 67). Cholesterol in the brain is involved in the formation of 'lipid rafts' which are implicat-

ed in cell transduction (68), and it also influences the activity of various enzymes, such as β -secretase (BACE1), which is involved in the metabolism of APP (67). The high concentration of cholesterol in these lipid rafts could facilitate the clustering of α - and β -secretases with their substrates in a configuration that leads to pathogenic cleavage of APP (67, 69). Additionally, a cluster of polymorphisms in cholesterol-related genes such as *APOE*, *SOAT1*, *APOE5'*-untranslated region, *OLR1*, *CYP46A1*, *LPL*, *LIPA* and *APOA4* has been shown to confer significant susceptibility to AD (70). Thus, a change in neuronal lipoprotein activity and cholesterol homeostasis poses an increased risk for the development of AD.

Excitotoxicity

Excitotoxicity describes the general phenomenon of neuronal cell death due to NMDA receptor-mediated calcium influx (71). It is theorized that excitotoxicity resulting from excessive activation of these receptors increases the vulnerability of CNS neurons, leading to neuronal degeneration (72) and causing synaptic failure (73). Recent studies have also shown that glutamine synthetase, an enzyme involved in the conversion of glutamate to glutamine, is oxidized in the brain of AD patients (74). Excessive glutamate in the synapse results in altered calcium homeostasis, excessive production of $A\beta_{42}$ and free radical generation, which ultimately leads to neuronal death.

Cholinergic deficit

AD is characterized by a deficit in cholinergic neurotransmission due to loss of cholinergic neurons of the

basal nucleus of Meynert (75). It has also been shown that the enzymes involved in the synthesis (choline acetyltransferase) and degradation (acetylcholinesterase) of acetylcholine have reduced activity and could be responsible for the deficit (76, 77). The basal forebrain cholinergic neurons are involved in the regulation of amyloidogenesis and hence APP processing pathways. A β peptide and NFTs cause neurotoxicity to the cholinergic innervations of the hippocampus and thus impair acetylcholine processing, leading to cognitive impairment.

Endocrine function

Menopause and andropause are characterized by a dramatic decline in sex steroids, resulting in an increase in the production of gonadotropins (78). It has been hypothesized that the marked reduction in sex hormone levels during the postmenopausal state results in a higher predisposition to develop AD (79, 80). Estrogen, which has been shown to have a neuroprotective effect by lowering brain A β levels (81) and improving synaptic plasticity (82), is depleted after menopause. This increases the risk for

AD. Luteinizing hormone (LH), a powerful endogenous mitogen, is also increased during aging and AD. This leads to dysregulation of the cell cycle in hippocampal neurons and results in initiation of the pathophysiological cascade of AD (83). Thus, hormonal changes associated with age also play a role in the etiopathogenesis of AD.

Therapeutic interventions

The development of a comprehensive therapeutic treatment for AD is limited by our understanding of the underlying biochemical mechanisms that cause neuronal failure. Numerous dysfunctional mechanisms have been described, ranging from protein aggregation and oxidative stress to biometal dyshomeostasis and mitochondrial failure (84). Thus, developing an effective therapeutic strategy for AD poses a great challenge for the scientific community. Numerous strategies have been developed and are under clinical evaluation, targeting different pathophysiological cascades of AD. A brief summary of the major pathophysiological events and the therapeutic targets are as shown in Figure 5.

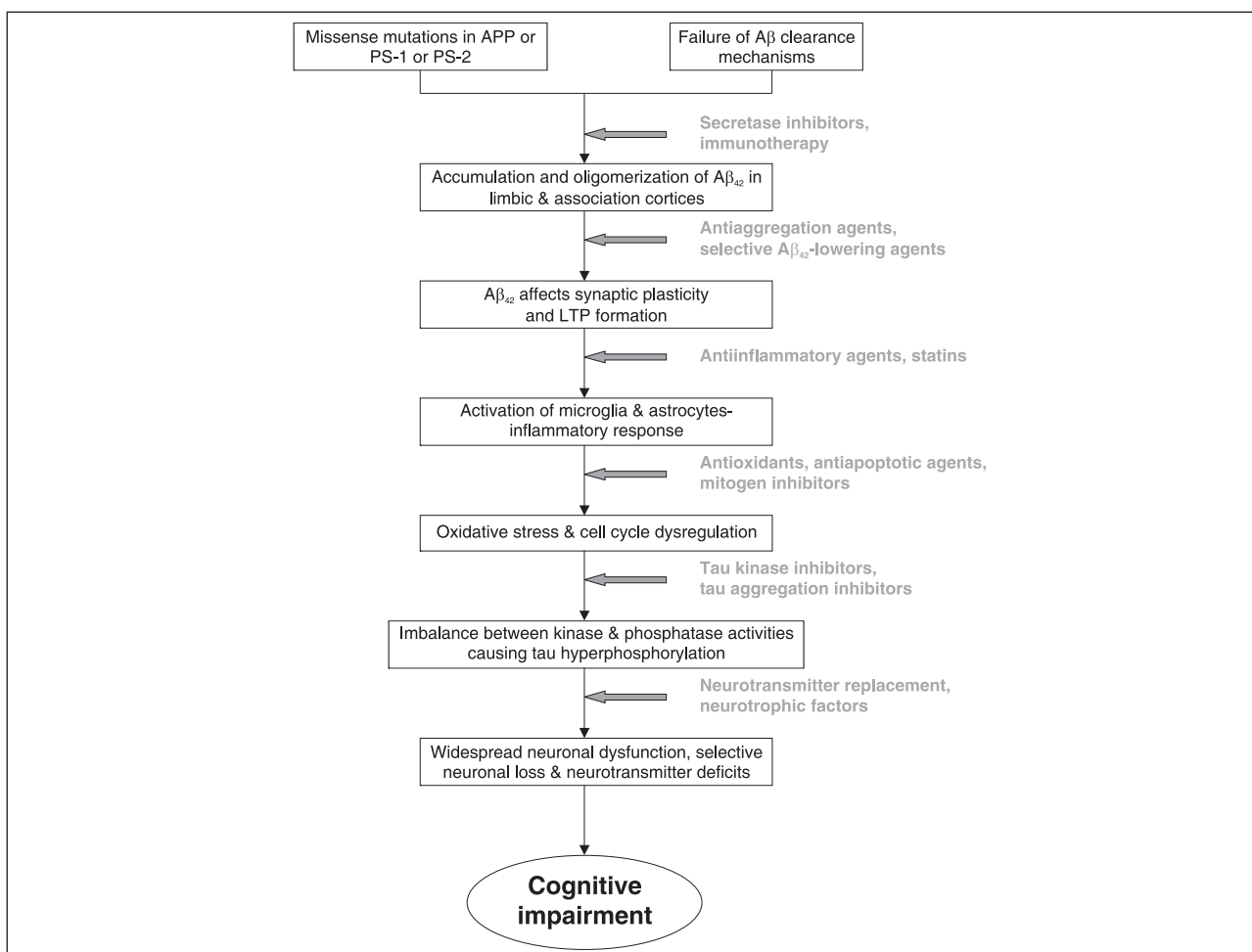


Fig. 5. A brief summary of the pathological events occurring in AD leading to cognitive impairment and selected strategies for therapeutic intervention.

Current FDA-approved drugs for AD and their limitations

AChE inhibitors (donepezil, rivastigmine and galantamine) and NMDA receptor antagonists (memantine) are the two classes of drugs approved for the management of AD. Both donepezil and galantamine are selective AChE inhibitors (85, 86), whereas rivastigmine is an inhibitor of both AChE and butyrylcholinesterase (BChE) (87). The cholinesterase inhibitors do exhibit improvements in patient memory and global function. However, their frequent use is limited by a short duration of action and an inadequate effect on disease progression (88). Memantine, developed on the basis of the excitotoxicity hypothesis, is another drug that has been approved for the symptomatic treatment of moderate to severe AD (89). It is only effective as a symptomatic agent, but does not halt disease progression. Thus, there is an urgent need to develop novel therapeutic strategies with strong disease-modifying properties.

Novel therapeutic strategies

1. Immunotherapy

Experiments proved that immunization with A β attenuates the AD-like pathology in a transgenic mouse model of AD (90). Active and passive immunization against A β can reduce learning deficits in APP transgenic mice (91, 92). The proposed mechanisms are that A β immunization triggers Fc receptor-mediated phagocytosis (90, 93) or can act as a chaperone and disrupt A β aggregates or prevent aggregation (94) and/or sequester A β , thereby shifting the equilibrium towards the periphery and reducing brain A β deposition (95). However, the main adverse effect of active immunization is aseptic meningioencephalitis (96-98). Therefore, alternative approaches need to be developed to avoid an unwanted T-cell response. Passive A β immunotherapy with monoclonal antibodies is also being evaluated, as well as DNA vaccines expressing A β and its fragments (17). Immunotherapy therefore holds great promise in future AD therapy.

2. Secretase inhibitors

The proteases, particularly γ -secretase and β -secretase, are attractive targets for developing drugs which could be used in the management of AD. Inhibitors of these secretases would block the formation of A β_{42} and prevent the subsequent downstream pathology (99). However, safety and pharmacokinetic problems have hindered drug development (100). LY-450139 (1) is a γ -secretase inhibitor that is presently in phase II clinical trials. It has been shown to reduce the cerebrospinal fluid (CSF) concentrations of A β . It is well tolerated and only a few cases of diarrhea have been reported (101). β -Secretase inhibitors that have been identified to date are large molecules and do not penetrate the blood-brain barrier (100, 102), and none of these molecules have progressed to clinical trials so far (103).

3. Selective A β_{42} -lowering agents

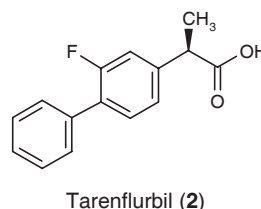
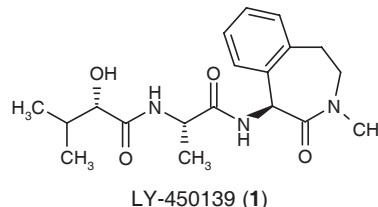
Tarenflurbil (2) is a pure (*R*)-enantiomer of flurbiprofen, the first in a novel class of disease-modifying drugs for mild AD (104, 105). This drug modulates γ -secretase, shifting the production away from A β_{42} towards the less toxic, shorter fragments of A β (A β_{38}). It also does not interfere with the function of Notch or other γ -secretase substrates (105). Several FDA-approved nonsteroidal antiinflammatory drugs (NSAIDs), such as ibuprofen, sulindac and indomethacin, have been shown to have selective A β_{42} -lowering properties. Although the exact mechanism is not clear, it is independent of cyclooxygenase inhibition (106). Thus, this therapeutic strategy is worthy of future investigation.

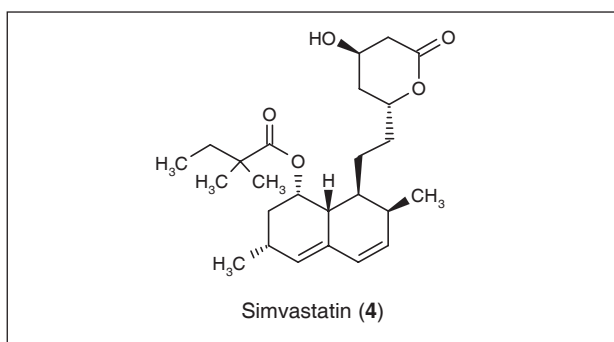
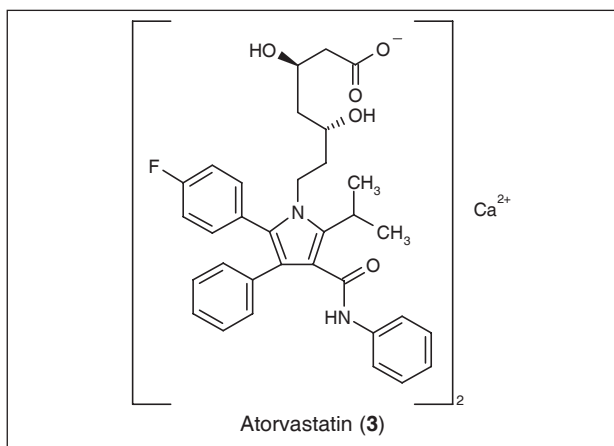
4. Statins

Epidemiological studies show that cholesterol-lowering drugs may have an impact on the progression of AD. The involvement of APOE, an established AD risk factor, in cholesterol metabolism provides support for this hypothesis (107). Statins act by preventing cholesterol-dependent cerebrovascular damage by inhibiting HMG-CoA reductase (108), and they upregulate α -secretase activity by inhibiting Rho-associated protein kinase 1 (ROCK1), an enzyme that modulates α -secretase activity. By enhancing the activity of α -secretase, the production of A β_{42} is prevented (109). Atorvastatin (3) and simvastatin (4) are presently in clinical trials.

5. Peroxisome proliferator-activated receptor γ (PPAR γ) agonists

The PPAR γ agonists rosiglitazone (5) and pioglitazone (6) are being assessed in phase II clinical trials as disease-modifying treatments for AD. These drugs act by reducing A β levels, plaque deposition and microglia-mediated inflammation (110). More recently, it has been demonstrated that PPAR γ regulates BACE1 mRNA, protein and activities via a PPAR response element (PPRE) in the promoter region of the BACE1 gene (111). It also modulates BACE1 activity at the molecular level, thereby influencing





APP metabolism. Apart from the glitazones, NSAIDs such as ibuprofen, indomethacin and naproxen also activate PPAR γ receptors and decrease the risk of AD (112, 113).

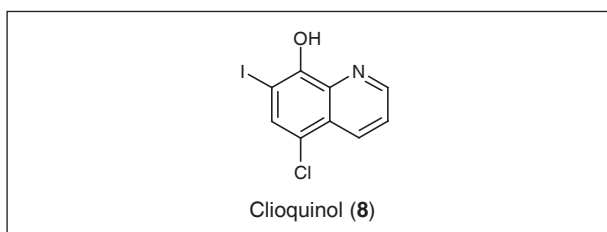
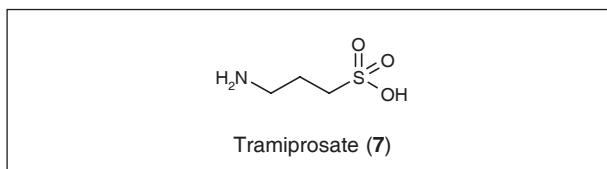
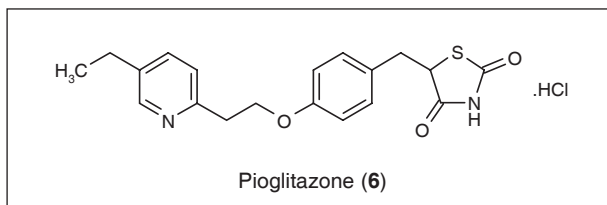
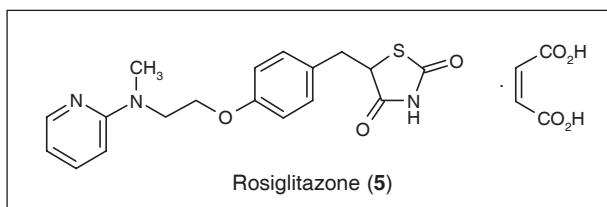
6. Antiaggregation agents

Preventing the formation of toxic oligomeric aggregates of A β using small molecules represents another approach for the development of novel therapeutics for treating AD. The glycosaminoglycan mimetic tramiprosate (7) is presently in phase III clinical trials (102). It reduces the plaque burden by competing with the sulfated glycosaminoglycans for the GAG binding sites and prevents fibril formation (114-116).

Another antiaggregant is O-CLN (ColostrininTM), a polypeptide complex derived from sheep colon that inhibits A β aggregation (117) and improves cognitive performance in animal models of AD (118). Following clinical trials in AD (119, 120), the company launched the product in 2007. Clioquinol (8), an antibiotic and Cu/Zn chelator, has also been shown to prevent aggregation of A β by chelating Zn and other divalent cations required for A β aggregation (121, 122). It is a reasonably well-tolerated drug in humans and is in phase II clinical trials for AD.

7. NSAIDs

Epidemiological evidence suggests that long-term use of NSAIDs protects against AD (123), but prospective studies of rofecoxib, naproxen and diclofenac failed to show their efficacy in delaying cognitive decline (124-126). Indomethacin has been shown to cross the blood-



brain barrier and improve cognitive function. This effect may be mediated by its ability to activate PPAR γ receptors, thereby inhibiting the activation of NF- κ B, and prevent the expression of inducible nitric oxide synthase (iNOS) and other inflammatory cytokines, such as tumor necrosis factor α (TNF- α) and IL-1 β (56). NSAIDs also have an allosteric effect on γ -secretase and thus reduce A β_{42} levels. However, this A β -lowering activity is achieved at high concentrations. These agents have therapeutic potential in AD, but their gastrointestinal and cardiovascular toxicity limits their use (127).

8. Antioxidants

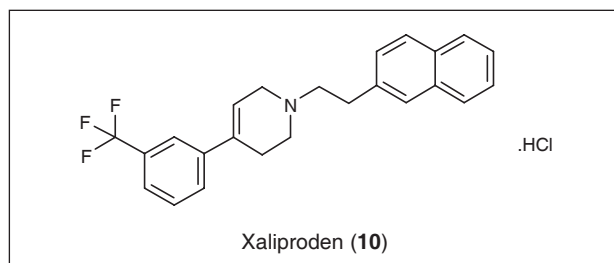
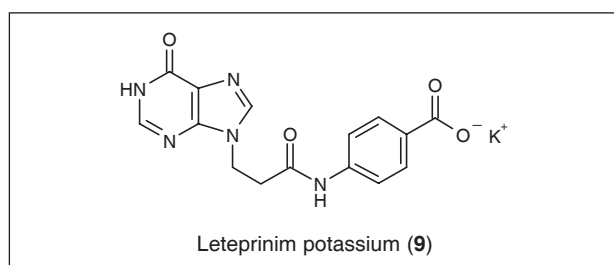
Oxidative damage appears to occur as one of the earliest pathophysiological events in AD and an increased intake of antioxidants lowers the risk of disease progression (128). Antioxidants such as acetyl-L-carnitine, aged garlic extract, α -lipoic acid, *Bacopa monniera*, ubiquinone, curcumin, ferulic acid, *Ginkgo biloba* extract, ginseng, green tea, huperzine A, melatonin, resveratrol, vitamin C and vitamin E have potential therapeutic value in AD (129). Their mechanism of action in AD management is shown in Table I.

9. Neurotrophic factors

NGF has been shown to stimulate the growth of cholinergic neurons, but the design of an effective deliv-

Table I: Brief summary of some common antioxidants and their proposed mechanism of action.

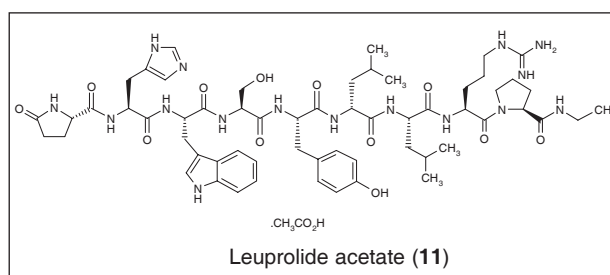
Antioxidant	Proposed mechanism of action in AD	Outcome in clinical trials
Acetyl-L-carnitine	Cholinomimetic, antioxidant	Promising
α -Lipoic acid	Mitochondrial antioxidant, neuroprotective	Promising
<i>Bacopa monniera</i>	Adaptogen, acetylcholinesterase inhibitor	Promising
Curcumin	Metal chelator, antioxidant, antiinflammatory, antiamyloidogenic	Promising
<i>Ginkgo biloba</i> extract	Antioxidant, antiapoptotic, neuroprotective against hippocampal cells, mitochondrial protector	Promising
Huperzine A	Antioxidant, reversible acetylcholinesterase inhibitor, memory enhancer	Promising
Melatonin	Antioxidant, antiamyloidogenic	Promising
Vitamin C & E (combination)	Antioxidants (chain terminators), neuroprotective	Promising



ery system is a challenge as NGF does not cross the blood-brain barrier (130). Leteprinin potassium (AIT-082; **9**) has been tested in a phase I study and was shown to increase the availability of NGF (131). Other related compounds, including xaliproden (SR-57746A; **10**; phase III), are also in clinical development.

10. Endocrine-based interventions

Dysregulation of reproductive hormones has been associated with the etiopathogenesis of AD. Gonadal hormones have been shown to be neuroprotective. 17 β -Estradiol and progesterone have been shown to possess antioxidant, antiinflammatory and antiamyloidogenic activity (132). Leuprolide acetate (**11**), an antigonadotropin, has entered phase III clinical trials for mild to moderate AD. The rationale for studying leuprolide is that LH is implicated in the pathogenesis of AD (133) and it has been shown to decrease the levels of A β ₄₂ in the brain tissue of mice (134). Given the diverse actions of reproductive hormones and their negative effects (increased risk of stroke, pulmonary embolism, etc.), further studies are needed.



11. Targeting tau hyperphosphorylation

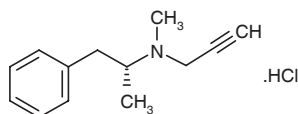
The phosphorylation of tau is controlled by different kinases and phosphatases. Protein phosphatase-2A (phosphoprotein phosphatase) increases the dephosphorylation of tau and also stimulates mitogen-activated protein kinases (MAPKs), which phosphorylate tau (135). Cyclin-dependent kinase 5 (CDK5) is another kinase which phosphorylates tau, and inhibitors of this enzyme may suppress tau phosphorylation and prevent tangle formation (136). Glycogen synthase kinase-3- β (GSK-3 β) has also been suggested as a drug target to inhibit tangle formation (137). Lithium, a mood stabilizer, has been shown to block this kinase, but further studies are required to prove its protective effect.

12. Monoamine oxidase (MAO) inhibitors

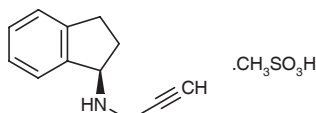
MAO inhibitors have neuroprotective effects. They have been shown to reduce the formation of toxic metabolites and ROS by blocking the enzymatic activity of MAO-A and MAO-B. Selegiline (**12**) has been shown to exert some symptomatic benefits in clinical trials in AD patients (138). Rasagiline (**13**; phase II) acts by influencing amyloid metabolism and induces antiapoptotic genes such as *BCL2* and *BCL-XL* (139, 140). They also interact with histone-modifying enzymes in the nucleus at the molecular level, thereby promoting gene expression (141).

13. Neurotransmitter replacement therapy

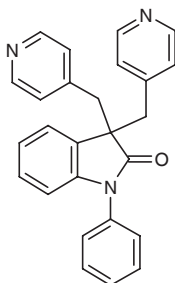
Acetylcholine has been thought of as the core neurotransmitter deficient in AD (142), but increasing evidence now suggests that noradrenergic (143), serotonergic (144, 145), γ -aminobutyric acid (GABA) (146) and



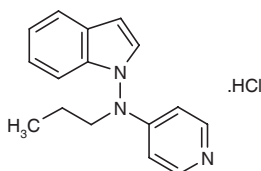
Selegiline (12)



Rasagiline (13)



Linopirdine (14)



Besipirdine (15)

dopaminergic system deficiencies may also be involved in the pathophysiological process of AD. Although numerous neurotransmitter replacement strategies have been attempted, none have been successful. One approach is now to administer agents that cause multisystem augmentation. Of particular interest are DuP-996 (linopirdine; **14**), an agent with positive effects on the cholinergic, serotonergic and dopaminergic systems, and HP-749 (besipirdine; **15**), which strengthens both cholinergic and noradrenergic systems. This approach seems promising and could have a role in the management of AD as adjunct therapy (147).

Conclusions

Given the current momentum of clinical research and developments in the understanding of the neuropathology of AD, improvements in the treatment of AD appear to be within sight. The focus will now be to diagnose the disease at an early stage so that novel disease-modifying

interventions can be used to achieve optimal benefit. Thus, as we enter the second century of AD, there is hope that improved treatment and diagnostic methods will soon be available.

Acknowledgements

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References

1. LaFerla, F.M., Green, K.N., Oddo, S. *Intracellular amyloid-beta in Alzheimer's disease*. Nat Rev Neurosci 2007, 8(7): 499-509.
2. Smith, M.A. *Alzheimer disease*. Int Rev Neurobiol 1998, 42: 1-54.
3. Robert, P.H., Verhey, F.R.J., Byrne, E.J. et al. *Grouping for behavioral and psychological symptoms in dementia: Clinical and biological aspects. Consensus paper of the European Alzheimer Disease Consortium*. Eur Psychiatry 2005, 20(7): 490-6.
4. Fuentealba, R.A., Farias, G., Scheu, J., Bronfman, M., Marzolo, M.P., Inestrosa, N.C. *Signal transduction during amyloid-beta peptide neurotoxicity: Role in Alzheimer's disease*. Brain Res Brain Res Rev 2004, 47(1-3): 275-89.
5. Alzheimer, A. *Über eine eigenartige Erkrankung der Hirnrinde*. Allgemeine Zeitschrift für Psychiatrie und Psychisch-Gerichtliche Medizin 1907, 64: 146-8.
6. Glenner, G.G., Wong, C.W. *Alzheimer's disease: Initial report of the purification and characterization of a novel cerebrovascular amyloid protein*. Biochem Biophys Res Commun 1984, 120(3): 885-90.
7. Masters, C.L. *Amyloid plaque core protein in Alzheimer's disease and Down syndrome*. Proc Natl Acad Sci USA 1985, 82(12): 4245-9.
8. Goedert, M., Wischik, C.M., Crowther, R.A., Walker, J.E., Klug, A. *Cloning and sequencing of the cDNA encoding core protein of the paired helical filament of Alzheimer disease: Identification as the microtubule-associated protein tau*. Proc Natl Acad Sci USA 1988, 85(11): 4051-5.
9. Grundke-Iqbal, I., Iqbal, K., Tung, Y.C., Quinlan, M., Wisniewski, H.M., Binder, L.I. *Abnormal phosphorylation of the microtubule-associated protein tau (tau) in Alzheimer cytoskeletal pathology*. Proc Natl Acad Sci USA 1986, 83(13): 4913-7.
10. Wyss-Coray, T. *Inflammation in Alzheimer disease: Driving force, bystander or beneficial response?* Nat Med 2006, 12(9): 1005-15.
11. Markesbery, W.R. *Oxidative stress hypothesis in Alzheimer's disease*. Free Radic Biol Med 1997, 23(1): 134-47.
12. Hebert, L.E., Scherr, P.A., Bienias, J.L., Bennett, D.A., Evans, D.A. *Alzheimer disease in the US population: Prevalence estimates using 2000 census*. Arch Neurol 2003, 60(8): 1119-22.
13. Ernst, R.L., Hay, J.W. *The US economic and social costs of Alzheimer's disease revisited*. Am J Public Health 1994, 84(8): 1261-4.

14. Clark, C.M., Karlawiah, J.H. *Alzheimer disease: Current concepts and emerging diagnostics and therapeutic strategies*. Ann Intern Med 2003, 138(5): 400-10.
15. Cummings, J.L. *Alzheimer's disease*. N Engl J Med 2004, 351(1): 56-67.
16. Hua, X., Lei, M., Zhang, Y., Ding, J., Han, Q., Hu, G., Xiao, M. *Long term D-galactose injection combined with ovariectomy serves as a new rodent model for Alzheimer's disease*. Life Sci 2007, 80(20): 1897-905.
17. Klafki, H.W., Staufenbiel, M., Kornhuber, J., Wiltfang, J. *Therapeutic approaches to Alzheimer's disease*. Brain 2006, 129(Pt. 11): 2840-55.
18. Hardy, J., Selkoe, D.J. *The amyloid hypothesis of Alzheimer's disease: Progress and problems on the road to therapeutics*. Science 2002, 297(5580): 353-6.
19. Lambert, M.P., Barlow, A.K., Chromy, B.A. et al. *Diffusible, nonfibrillar ligands derived from Abeta1-42 are potent central nervous system neurotoxins*. Proc Natl Acad Sci USA 1998, 95(11): 6448-53.
20. Hartley, D.M., Walsh, D.M., Ye, C.P. et al. *Protofibrillar intermediates of amyloid beta-protein induce acute electrophysiological changes and progressive neurotoxicity in cortical neurons*. J Neurosci 1999, 19(20): 8876-84.
21. Weidemann, A., König, G., Bunke, D., Fischer, P., Salbaum, J.M., Masters, C.L., Beyreuther, K. *Identification, biogenesis, and localization of precursors of Alzheimer's disease A4 amyloid protein*. Cell 1989, 57(1): 115-26.
22. Esch, F.S., Keim, P.S., Beattie, E.C. et al. *Cleavage of amyloid beta peptide during constitutive processing of its precursor*. Science 1990, 248(4959): 1122-4.
23. Sisodia, S., Koo, E., Beyreuther, K., Unterbeck, A., Price, D.L. *Evidence that beta-amyloid protein in Alzheimer's disease is not derived by normal processing*. Science 1990, 248(4954): 492-5.
24. Verdile, G., Fuller, S., Atwood, C.S., Laws, S.M., Gandy, S.E., Martins, R.N. *The role of beta amyloid in Alzheimer's disease: Still a cause of everything or only one who got caught?* Pharmacol Res 2004, 50(4): 397-409.
25. Reddy, P.H. *Amyloid precursor protein-mediated free radicals and oxidative damage: Implications for the development and progression of Alzheimer's disease*. J Neurochem 2006, 96(1): 1-13.
26. Rottkamp, C.A., Atwood, C.S., Joseph, J.A., Nunomura, A., Perry, G., Smith, M.A. *The state versus amyloid-beta: The trial of the most wanted criminal in Alzheimer disease*. Peptides 2002, 23(7): 1333-41.
27. Smith, M.A., Casadesus, G., Joseph, J.A., Perry, G. *Amyloid-beta and tau serve antioxidant functions in the aging and Alzheimer brain*. Free Radic Biol Med 2002, 33(9): 1194-9.
28. Roder, H.M., Hutton, M.L. *Microtubule-associated protein tau as a therapeutic target in neurodegenerative disease*. Expert Opin Ther Targets 2007, 11(4): 435-42.
29. Heston, L.L., White, J. *Pedigrees of 30 families with Alzheimer's disease: Associations with defective organization of microfilaments and microtubules*. Behav Genet 1978, 8(4): 315-31.
30. Paula-Barbosa, M., Tavares, M.A., Cadete-Leite, A. *A quantitative study of frontal cortex dendritic microtubules in patients with Alzheimer's disease*. Brain Res 1987, 417(1): 139-42.
31. Yoshida, H., Ihara, Y. *Tau in paired helical filaments is functionally distinct from fetal tau: Assembly incompetence of paired helical filament-tau*. J Neurochem 1993, 61(3): 1183-6.
32. Spires, T.L., Orne, J.D., Santacruz, K., Pitstick, R., Carlson, G.A., Ashe, K.H., Hyman, B.T. *Region specific dissociation of neuronal loss and neurofibrillary pathology in a mouse model of tauopathy*. Am J Pathol 2006, 168(5): 1598-607.
33. Andorfer, C., Acker, C.M., Kress, Y., Hof, P.R., Duff, K., Davies, P. *Cell-cycle reentry and cell death in transgenic mice expressing nonmutant human tau isoforms*. J Neurosci 2005, 25(22): 5446-54.
34. Moreira, P.I., Zhu, X., Liu, Q. et al. *Compensatory responses induced by oxidative stress in Alzheimer disease*. Biol Res 2006, 39(1): 7-13.
35. Cross, C.E., Halliwell, B., Borish, W.A. et al. *Oxygen radicals and human disease*. Ann Intern Med 1987, 107(4): 526-45.
36. Smith, M.A., Sayre, L.M., Monnier, V.M., Perry, G. *Radical AGEing in Alzheimer's disease*. Trends Neurosci 1995, 18(4): 172-6.
37. Mecocci, P., MacGarvey, U., Beal, M.F. *Oxidative damage to mitochondrial DNA is increased in Alzheimer's disease*. Ann Neurol 1994, 36(5): 747-51.
38. Nunomura, A., Perry, G., Pappolla, R., Wade, R., Hirai, K., Chiba, S., Smith, M.A. *RNA oxidation is a prominent feature of vulnerable neurons in Alzheimer's disease*. J Neurosci 1999, 19(6): 1959-64.
39. Perry, G., Taddeo, M.A., Petersen, R.B. et al. *Adventitiously bound redox active iron and copper are at the center of oxidative damage in Alzheimer's disease*. Biometals 2003, 16(1): 77-81.
40. Butterfield, D.A., Drake, J., Pocernich, C., Castegna, A. *Evidence of oxidative damage in Alzheimer's disease brain: Central role for amyloid beta-peptide*. Trends Mol Med 2001, 7(12): 548-54.
41. Smith, M.A., Kutty, R.K., Richey, P.L. et al. *Heme oxygenase-1 is associated with neurofibrillary pathology of Alzheimer's disease*. Am J Pathol 1994, 145(1): 42-7.
42. Zhu, X., Webber, K.M., Casadesus, G. et al. *Mitotic and gender parallels in Alzheimer's disease: Therapeutic opportunities*. Curr Drug Targets 2004, 5(6): 559-63.
43. Bowser, R., Smith, M.A. *Cell cycle proteins in Alzheimer's disease: Plenty of wheels but no cycle*. J Alzheimers Dis 2002, 4(3): 249-54.
44. Hirai, K., Aliev, G., Nunomura, A. et al. *Mitochondrial abnormalities in Alzheimer's disease*. J Neurosci 2001, 21(9): 3017-23.
45. Jordan-Sciutto, K.L., Malaiyandi, L.M., Bowser, R. *Altered distribution of cell cycle transcriptional regulators during Alzheimer's disease*. J Neuropathol Exp Neurol 2002, 61(4): 358-67.
46. Tsujioka, Y., Takahashi, M., Tsuboi, Y., Yamamoto, T., Yamada, T. *Localization and expression of cdc2 and cdk4 in Alzheimer brain tissue*. Dement Geriatr Cogn Disord 1999, 10(3): 192-8.

47. Khurana, V., Lu, Y., Steinhilb, M.L., Oldham, S., Shulman, J.M., Feany, M.B. *TOR-mediated cell cycle activation causes neurodegeneration in Drosophila tauopathy model*. *Curr Biol* 2006, 16(3): 230-41.
48. Kalara, R.N., Gravina, S.A., Schmidley, J.W., Perry, G., Harik, S.I. *The glucose transporter of the human brain and blood-brain barrier*. *Ann Neurol* 1988, 24(6): 757-64.
49. Zhu, X., Raina, A.K., Lee, H., Casadesus, G., Smith, M.A., Perry, G. *Oxidative stress signaling in Alzheimer's disease*. *Brain Res* 2004, 1000(1-2): 32-9.
50. Verdile, G., Gandy, S.E., Martins, R.N. *The role of presenilin and interacting proteins in the biogenesis of Alzheimer's beta amyloid*. *Neurochem Res* 2007, 32(4-5): 609-23.
51. Borchelt, D.R., Thinakaran, G., Eckman, C.B. et al. *Familial Alzheimer's disease-linked presenilin 1 variants elevate Abeta1-42/1-40 ratio in vitro and in vivo*. *Neuron* 1996, 17(5): 1005-13.
52. Lemere, C.A., Lopera, F., Kosik, K.S. et al. *The E280A presenilin 1 Alzheimer mutation produces increased Abeta42 deposition and severe cerebellar pathology*. *Nat Med* 1996, 2(10): 1146-50.
53. Xia, W., Zhang, J., Kholodenko, D. et al. *Enhanced production and oligomerization of the 42-residue amyloid beta-protein by Chinese hamster ovary cells stably expressing mutant presenilins*. *J Biol Chem* 1997, 272(12): 7977-82.
54. Nilsberth, C., Westlind-Danielsson, A., Eckman, C.B. et al. *The 'Arctic' APP mutation (E693G) causes Alzheimer's disease by enhanced Abeta protofibril formation*. *Nat Neurosci* 2001, 4(9): 887-93.
55. Scheuner, D., Eckman, C., Jensen, M. et al. *Secreted amyloid beta-proteins similar to that in the senile plaques of Alzheimer's disease is increased in vivo by the presenilin 1 and 2 and APP mutations linked to familial Alzheimer's disease*. *Nat Med* 1996, 2(8): 864-70.
56. Hoozemans, J.J.M., O'Banion, M.K. *The role of COX-1 and COX-2 in Alzheimer's disease pathology and the therapeutic potentials of non-steroidal anti-inflammatory drugs*. *Curr Drug Targets CNS Neurol Disord* 2005, 4(3): 307-15.
57. McGeer, P.L., McGeer, E.G. *The inflammatory response system of brain: Implications for therapy of Alzheimer and other neurodegenerative diseases*. *Brain Res Brain Res Rev* 1995, 21(2): 195-218.
58. De Servi, B., La Porta, C.A., Bontempelli, M., Comolli, R. *Decrease of TGF-beta1 plasma levels and increase in nitric oxide synthase activity in leukocytes as potential biomarkers for Alzheimer's disease*. *Exp Gerontol* 2002, 37(6): 813-21.
59. Malaguarnera, L., Motta, M., Di Rosa, M., Anzaldi, M., Malaguarnera, M. *Interleukin-18 and transforming growth factor-beta1 plasma levels in Alzheimer's disease and vascular dementia*. *Neuropathology* 2006, 26(4): 307-12.
60. Saez, T.E., Pehar, M., Vargas, M., Barbeito, L., Maccioni, R.B. *Astrocytic nitric oxide triggers tau hyperphosphorylation in hippocampal neurons*. *In Vivo* 2004, 18(3): 275-80.
61. Saez, E.T., Pehar, M., Vargas, M., Barbeito, L., Maccioni, R.B. *Production of nerve growth factor by beta-amyloid-stimulated astrocytes induces p75NTR-dependent tau hyperphosphorylation in cultured hippocampal neurons*. *J Neurosci Res* 2006, 84(5): 1098-106.
62. Quintanilla, R.A., Orellana, D.I., González-Billault, C., Maccioni, R.B. *Interleukin-6 induces Alzheimer type phosphorylation of tau protein by deregulating the cdk5/p35 pathway*. *Exp Cell Res* 2004, 295(1): 245-57.
63. Campbell, I.L., Stadler, A.K., Atwa, Y., Pagenstecher, A., Asensio, V.C. *Transgenic models to study the actions of cytokines in the central nervous system*. *Neuroimmunomodulation* 1998, 5(3-4): 126-35.
64. Heyser, C.J., Masliah, E., Samimi, A., Campbell, I.L., Gold, L.H. *Progressive decline in avoidance learning paralleled by inflammatory neurodegeneration in transgenic mice expressing interleukin-6 in the brain*. *Proc Natl Acad Sci USA* 1997, 94(4): 1500-5.
65. Rojo, L.E., Fernández, J.A., Maccioni, A.A., Jimenez, J.M., Maccioni, R.B. *Neuroinflammation: Implications for the pathogenesis and molecular diagnosis of Alzheimer's disease*. *Arch Med Res* 2007, 39(1): 1-16.
66. Pappolla, M.A., Bryant-Thomas, T.K., Herbert, D. et al. *Mild hypercholesterolemia is an early risk factor for the development of Alzheimer amyloid pathology*. *Neurology* 2003, 61(2): 199-205.
67. Kuo, Y.-M., Emmerling, M.R., Bisgaier, C.L., Essenburg, A.D., Lampert, H.C., Drumm, D., Roher, A.E. *Elevated low-density lipoprotein in Alzheimer's disease correlates with brain Abeta 1-42 levels*. *Biochem Biophys Res Commun* 1998, 252(3): 711-5.
68. Rojo, L., Sjöberg, M.K., Hernández, P., Zambrano, C., Maccioni, R.B. *Roles of cholesterol and lipids in the etiopathogenesis of Alzheimer's disease*. *J Biomed Biotechnol* 2006, 2006(3): 73976.
69. Desire, L., Bourdain, J., Loiseau, N. et al. *RAC1 inhibition targets amyloid precursor protein processing by gamma-secretase and decreases Abeta production in vitro and in vivo*. *J Biol Chem* 2005, 280(45): 37516-25.
70. Papassotiropoulos, A., Wollmer, M.A., Tsolaki, M. et al. *A cluster of cholesterol-related genes confer susceptibility for Alzheimer's disease*. *J Clin Psychiatry* 2005, 66(7): 940-7.
71. Hüll, M., Berger, M., Heneka, M. *Disease-modifying therapies in Alzheimer's disease. How far have we come?* *Drugs* 2006, 66(16): 2075-93.
72. Castellani, R.J., Zhu, X., Lee, H.G., Moreira, P.I., Perry, G., Smith, M.A. *Neuropathology and treatment of Alzheimer disease: Did we lose the forest for the trees?* *Expert Rev Neurother* 2007, 7(5): 473-85.
73. Selkoe, D.J. *Alzheimer's disease is a synaptic failure*. *Science* 2002, 298(5594): 789-91.
74. Castegna, A., Aksenov, M., Aksenova, M. et al. *Proteomic identification of oxidatively modified proteins in Alzheimer's diseased brain. Part I: Creatine kinase BB, glutamine synthase, and ubiquitin carboxy-terminal hydrolase L-1*. *Free Radic Biol Med* 2002, 33(4): 562-71.
75. Kasa, P., Rakonczay, Z., Gulya, K. *The cholinergic system in Alzheimer's disease*. *Prog Neurobiol* 1997, 52(6): 511-35.
76. Sims, N.R., Bowen, D.M., Allen, S.J., Smith, C.C., Neary, D., Thomas, D.J., Davison, A.N. *Presynaptic cholinergic dysfunction in patients with dementia*. *J Neurochem* 1983, 40(2): 503-9.

77. DeKosky, S.T., Harbaugh, R.E., Schmitt, F.A. et al. *Cortical biopsy in Alzheimer's disease: Diagnostic accuracy and neurochemical, neuropathological and cognitive correlations. Intraventricular Bethanecol Study Group.* Ann Neurol 1992, 32(5): 625-32.
78. Casadesus, G., Puig, E.R., Webber, K.M. et al. *Targeting gonadotropins: An alternative option for Alzheimer disease treatment.* J Biomed Biotechnol 2006, 206(3): 39508.
79. Letenneur, L., Commenges, D., Dartigues, J.F., Barberger-Gateau, P. *Incidence of dementia and Alzheimer's disease in elderly community residents of south-western France.* Int J Epidemiol 1994, 23(6): 1256-61.
80. Rocca, W.A., Hofman, A., Brayne, C. et al. *Frequency and distribution of Alzheimer's disease in Europe: A collaborative study of 1980-1990 prevalence findings. The EURODEM-Prevalence Research Group.* Ann Neurol 1991, 30(3): 381-90.
81. Petanceska, S.S., Nagy, V., Frail, D., Gandy, S. *Ovariectomy and 17beta-estradiol modulate the levels of Alzheimer's amyloid beta peptides in brain.* Neurology 2000, 54(12): 2212-7.
82. Bi, R., Foy, M.R., Vouimba, R.M., Thompson, R.F., Baudry, M. *Cyclic changes in estradiol regulate synaptic plasticity through the MAP kinase pathway.* Proc Natl Acad Sci USA 2001, 98(23): 13391-5.
83. Zhu, X., Raina, A.K., Perry, G., Smith, M.A. *Alzheimer's disease: The two-hit hypothesis.* Lancet Neurol 2004, 3(4): 219-26.
84. Crouch, P.J., Harding, S.M.E., White, A.R., Camakaris, J., Bush, A.I., Masters, C.L. *Mechanisms of Abeta mediated neurodegeneration in Alzheimer's disease.* Int J Biochem Cell Biol 2008, 40(2): 181-98.
85. Kryger, G., Silman, I., Sussman, J.L. *Three-dimensional structure of a complex of E2020 with acetylcholinesterase from Torpedo californica.* J Physiol (Paris) 1998, 92(3-4): 191-4.
86. Greenblatt, H.M., Kryger, G., Lewis, T., Silman, I., Sussman, J.L. *Structure of acetylcholinesterase complexed with (-)-galanthamine at 2.3 Å resolution.* FEBS Lett 1999, 463(3): 321-6.
87. Farlow, M. *A clinical overview of cholinesterase inhibitors in Alzheimer's disease.* Int Psychogeriatrics 2002, 14(Suppl. 1): 93-126.
88. Jacobsen, J.S., Reinhart, P., Pangalos, M.N. *Current concepts in therapeutic strategies targeting cognitive decline and disease modification in Alzheimer's disease.* NeuroRx 2005, 2(4): 612-26.
89. Bullock, R. *New drugs for Alzheimer's disease and other dementias.* Br J Psychiatry 2002, 180(2): 135-9.
90. Schenk, D., Barbour, R., Dunn, W. et al. *Immunization with amyloid-beta attenuates Alzheimer-disease-like pathology in the PDAPP mouse.* Nature 1999, 400(6740): 173-7.
91. Janus, C., Pearson, J., McLaurin, J. et al. *Abeta peptide immunization reduces behavioral impairment and plaques in a model of Alzheimer's disease.* Nature 2000, 408(6815): 979-82.
92. Morgan, D., Diamond, D.M., Gottschall, P.E.J. et al. *Abeta peptide vaccination prevents memory loss in an animal model of Alzheimer's disease.* Nature 2000, 408(6815): 982-5.
93. Bard, F., Cannon, C., Barbour, R. et al. *Peripherally administered antibodies against amyloid beta-peptide enter the central nervous system and reduce pathology in a mouse model of Alzheimer's disease.* Nat Med 2000, 6(8): 916-9.
94. Solomon, B. *Generation of anti-beta amyloid antibodies via phage display technology towards Alzheimer's disease vaccination.* Vaccine 2005, 23(17-18): 2327-30.
95. DeMattos, R.B., Bales, K.R., Cummins, D.J., Dodart, J.C., Paul, S.M., Holtzman, D.M. *Peripheral anti-Abeta antibody alters CNS and plasma Abeta clearance and decreases brain Abeta burden in a mouse model of Alzheimer's disease.* Proc Natl Acad Sci USA 2001, 98(15): 8850-5.
96. Orgogozo, J.M., Gilman, S., Dartigues, J.F. et al. *Subacute meningoencephalitis in a subset of patients with AD after Abeta42 immunization.* Neurology 2003, 61(1): 46-54.
97. Bayer, A.J., Bullock, R., Jones, R.W. et al. *Evaluation of the safety and immunogenicity of synthetic Abeta42 (AN1792) in patients with AD.* Neurology 2005, 64(1): 94-101.
98. Gilman, S., Koller, M., Black, R.S. et al. *Clinical effects of Abeta immunization (AN1792) in patients with AD in an interrupted trial.* Neurology 2005, 64(9): 1553-62.
99. Christensen, D.D. *Alzheimer's disease: Progress in the development of anti-amyloid disease-modifying therapies.* CNS Spectr 2007, 12(2): 113-23.
100. Walker, L.C., Ibegbu, C.C., Todd, C.W., Robinson, H.L., Jucker, M., LeVine, H. 3rd, Gandy, S. *Emerging prospects for the disease-modifying treatment of Alzheimer's disease.* Biochem Pharmacol 2005, 69(7): 1001-8.
101. Siemers, E., Skinner, M., Dean, R.A. et al. *Safety, tolerability and changes in amyloid beta concentrations after administration of a gamma-secretase inhibitor in volunteers.* Clin Neuropharmacol 2005, 28(3): 126-32.
102. Citron, M. *Strategies for disease modification in Alzheimer's disease.* Nat Rev Neurosci 2004, 5(9): 677-85.
103. Rosenberg, R.N. *Translational research on the way to effective therapy for Alzheimer disease.* Arch Gen Psychiatry 2005, 62(11): 1186-92.
104. Golde, T.E. *Alzheimer disease therapy: Can the amyloid cascade be halted?* J Clin Invest 2003, 111(1): 11-8.
105. Weggen, S., Eriksen, J.L., Sagi, S.A., Pietrzik, C.U., Golde, T.E., Koo, E.H. *Abeta42-lowering nonsteroidal anti-inflammatory drugs preserve intramembrane cleavage of the amyloid precursor protein (APP) and ErbB-4 receptor and signaling through the APP intracellular domain.* J Biol Chem 2003, 278(33): 30748-54.
106. Weggen, S., Eriksen, J.L., Das, P. et al. *A subset of NSAIDs lower amyloidogenic Abeta42 independently of cyclooxygenase activity.* Nature 2001, 414(6860): 212-6.
107. Silvestrelli, G., Lanari, A., Parnetti, L., Tomassoni, D., Amenta, F. *Treatment of Alzheimer's disease: From pharmacology to a better understanding of disease pathophysiology.* Mech Ageing Dev 2006, 127(2): 148-57.
108. Whitfield, J.F. *Can statins put the brakes on Alzheimer's disease?* Expert Opin Investig Drugs 2006, 15(12): 1479-85.
109. Pedrini, S., Carter, T.L., Prendergast, G., Petanceska, S., Ehrlich, M.E., Gandy, S. *Modulation of statin-activated shedding of Alzheimer APP ectodomain by ROCK.* PLoS Med 2005, 2(1): e18.

110. Heneka, M.T., Sastre, M., Dumitrescu-Ozimek, L. et al. *Acute treatment with PPARgamma agonists pioglitazone and ibuprofen reduces glial inflammation and Abeta1-42 levels in APPV7171 transgenic mice.* Brain 2005, 128(Pt. 6): 1442-53.
111. Sastre, M., Klockgether, T., Heneka, M.T. *Contribution of inflammatory processes to Alzheimer's disease: Molecular mechanisms.* Int J Dev Neurosci 2006, 24(2-3): 167-76.
112. In't Veld, B.A., Ruitenbergh, A., Hofman, A. et al. *Nonsteroidal antiinflammatory drugs and the risk of Alzheimer's disease.* N Engl J Med 2001, 345(21): 1515-21.
113. Lehmann, J.M., Lenhard, J.M., Oliver, B.B., Ringold, G.M., Kliewer, S.A. *Peroxisome proliferator-activated receptors alpha and gamma are activated by indomethacin and other non-steroidal anti-inflammatory drugs.* J Biol Chem 1997, 272(6): 3406-10.
114. Gervais, F., Chalifour, R., Garceau, D. et al. *Glycosaminoglycan mimetics: A therapeutic approach to cerebral amyloid angiopathy.* Amyloid 2001, 8(Suppl. 1): 28-35.
115. Garceau, D., Gurbindo, C., Laurin, J. *Safety, tolerability and pharmacokinetic profile of Alzhemed, an anti-amyloid agent for Alzheimer's disease, in healthy subjects.* 7th Int Geneva/Springfield Symp Adv Alzheimer Ther (April 3-6, Geneva) 2002.
116. Geerts, H. *NC-531 (Neurochem).* Curr Opin Investig Drugs 2004, 5(1): 95-100.
117. Gibson, G.L., Douraghi-Zadeh, D., Parsons, R.B., Austen, B.M. *Properties of ovine colostrinin (O-CLN) on the in vitro aggregation and toxicity of beta-amyloid.* Neurobiol Aging 2004, 25: 592.
118. Rattray, M. *Technology evaluation: Colostrinin, ReGen.* Curr Opin Mol Ther 2005, 7(1): 78-84.
119. Bilikiewicz, A., Gaus, W. *Colostrinin (a naturally occurring, proline-rich, polypeptide mixture) in the treatment of Alzheimer's disease.* J Alzheimers Dis 2004, 6(1): 17-26.
120. Leszek, J., Inglot, A.D., Janusz, M., Byczkiewicz, F., Kiejna, A., Georgiades, J., Lisowski, J. *Colostrinin proline-rich polypeptide complex from ovine colostrums — A long-term study of its efficacy in Alzheimer's disease.* Med Sci Monit 2002, 8(10): PI193-6.
121. Cherny, R.A., Atwood, C.S., Xilinas, M.E. et al. *Treatment with copper-zinc chelator markedly and rapidly inhibits beta-amyloid accumulation in Alzheimer's disease transgenic mice.* Neuron 2001, 30(3): 665-76.
122. Curtain, C.C., Ali, F., Volitakis, I. et al. *Alzheimer's disease amyloid-beta binds copper and zinc to generate an allosterically ordered membrane-penetrating structure containing superoxide dismutase-like subunits.* J Biol Chem 2001, 276(23): 20466-73.
123. Szekely, C.A., Thorne, J.E., Zandi, P.P., Ek, M., Messias, E., Breitner, J.C., Goodman, S.N. *Nonsteroidal anti-inflammatory drugs for the prevention of Alzheimer's disease: A systematic review.* Neuroepidemiology 2004, 23(4): 159-69.
124. Aisen, P.S., Schafer, K.A., Grundman, M. et al. *Effects of rofecoxib or naproxen vs placebo on Alzheimer disease progression. A randomized controlled trial.* JAMA — J Am Med Assoc 2003, 289(21): 2819-26.
125. Reines, S.A., Block, G.A., Morris, J.C. et al. *Rofecoxib: No effect on Alzheimer's disease in a 1-year, randomized, blinded, controlled study.* Neurology 2004, 62(1): 66-71.
126. Scharf, S., Mander, A., Ugoni, A., Vajda, F., Christophidis, N. *A double-blind, placebo-controlled trial of diclofenac/misoprostol in Alzheimer's disease.* Neurology 1999, 53(1): 197-201.
127. Firuzi, O., Praticò, D. *Coxibs and Alzheimer's disease: Should they stay or should they go?* Ann Neurol 2006, 59(2): 219-28.
128. Frank, B., Gupta, S. *A review of antioxidants and Alzheimer's disease.* Ann Clin Psychiatry 2005, 17(4): 269-86.
129. Prasad, K.N., Cole, W.C., Prasad, K.C. *Risk factors for Alzheimer's disease: Role of multiple antioxidants, non-steroidal anti-inflammatory and cholinergic agents alone or in combination in prevention and treatment.* J Am Coll Nutr 2002, 21(6): 506-22.
130. Blesch, A., Tuszynski, M.H. *Gene therapy and cell transplantation for Alzheimer's disease and spinal cord injury.* Yonsei Med J 2004, 45(Suppl.): 28-31.
131. Grundman, M., Capparelli, E., Kim, H.T. et al. *A multicenter, randomized, placebo controlled, multiple-dose, safety and pharmacokinetic study of AIT-082 (Neotrofin) in mild Alzheimer's disease patients.* Life Sci 2003, 73(5): 539-53.
132. Barron, A.M., Fuller, S.J., Verdile, G., Martins, R.N. *Reproductive hormones modulate oxidative stress in Alzheimer's disease.* Antioxid Redox Signal 2006, 8(11-12): 2047-59.
133. Bowen, R.L., Smith, M.A., Harris, P.L. et al. *Elevated luteinizing hormone expression colocalizes with neurons vulnerable to Alzheimer's disease pathology.* J Neurosci Res 2002, 70(3): 514-8.
134. Bowen, R.L., Verdile, G., Liu, T.B. et al. *Luteinizing hormone, a reproductive regulator that modulates the processing of amyloid-beta precursor protein and amyloid-beta deposition.* J Biol Chem 2004, 279(19): 20539-45.
135. Iqbal, I., Grundke-Iqbal, I. *Pharmacological approaches of neurofibrillary degeneration.* Curr Alzheimer Res 2005, 2(3): 335-41.
136. Lau, L.F., Seymour, P.A., Sanner, M.A., Schachter, J.B. *Cdk5 as a drug target for the treatment of Alzheimer's disease.* J Mol Neurosci 2002, 19(3): 267-73.
137. Bhat, R.V., Budd Haeberlein, S.L., Avila, J. *Glycogen synthase kinase 3: A drug target for CNS therapies.* J Neurochem 2004, 89(6): 1313-7.
138. Birks, J., Flicker, L. *Selegiline for Alzheimer's disease.* Cochrane Database Syst Rev 2003, 1: CD000442.
139. Youdim, M.B., Fridkin, M., Zheng, H. *Bifunctional drug derivatives of MAO-B inhibitor rasagiline and iron chelator VK-28 as a more effective approach to treatment of brain ageing and ageing neurodegenerative diseases.* Mech Ageing Dev 2005, 126(2): 317-26.
140. Youdim, M.B., Weinstock, M. *Molecular basis of neuroprotective activities of rasagiline and the anti-Alzheimer drug TV3326 [(N-propargyl-(3R)aminoindan-5-yl)-ethyl methyl carbamate].* Cell Mol Neurobiol 2001, 21(6): 555-73.
141. Metzger, E., Wissmann, M., Yin, N. et al. *LSD1 demethylates repressive histone marks to promote androgen-receptor-dependent transcription.* Nature 2005, 437(7057): 436-9.

142. Perry, E.K., Tomlinson, B.E., Blessed, G., Bergmann, K., Gibson, B.E., Perry, R.H. *Correlation of cholinergic abnormalities with senile plaques and mental test scores in senile dementia*. Br Med J 1978, 2(6150): 1457-9.
143. Narang, P.K., Cutlet, N.R. *Pharmacotherapy in Alzheimer's disease: Basis and rationale*. Prog Neuropsychopharmacol Biol Psychiatry 1986, 10(3-5): 519-31.
144. Whitford, G.M. *Alzheimer's disease and serotonin: A review*. Neuropsychobiology 1986, 15(3-4): 133-42.
145. Palmer, A.M., Francis, P.T., Benton, J.S. et al. *Presynaptic serotonergic dysfunction in patients with Alzheimer's disease*. J Neurochem 1987, 48(1): 8-15.
146. Coyle, J.T., Price, D.L., DeLong, M.R. *Alzheimer's disease: A disorder of cortical cholinergic innervation*. Science 1983, 219(4589): 1184-90.
147. Mohr, E., Mendis, T., Rusk, I.N., Grimes, J.D. *Neurotransmitter replacement therapy in Alzheimer's disease*. J Psychiatry Neurosci 1994, 19(1): 17-23.